

pUC19 DNA/Mspl (Hpall) Marker, 23

#SM0221 50 μg

(for 100 applications)

Lot:

Concentration: 0.5 µg/µL

Supplied with: 1 mL 6X DNA Loading Dye

Store at -20°C

In total 2 vials.

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Description

pUC19 DNA was completely digested with Mspl, purified and dissolved in a storage buffer.

The DNA Marker contains the following 13 discrete fragments (in base pairs): 501, 489, 404, 331, 242, 190, 147, 111, 110, 67, 34, 34, 26.

Storage Buffer

10 mM Tris-HCl (pH 7.6), 1 mM EDTA.

6X DNA Loading Dye

10 mM Tris-HCl (pH 7.6), 0.03% bromophenol blue, 0.03% xylene cyanol FF, 60% glycerol and 60 mM EDTA.

CERTIFICATE OF ANALYSIS

Well-defined bands are formed during agarose gel electrophoresis.

The DNA concentration is determined spectrophptometrically.

The absence of nucleases is confirmed by a direct nuclease activity assay.

Quality authorized by:



Jurgita Zilinskiene

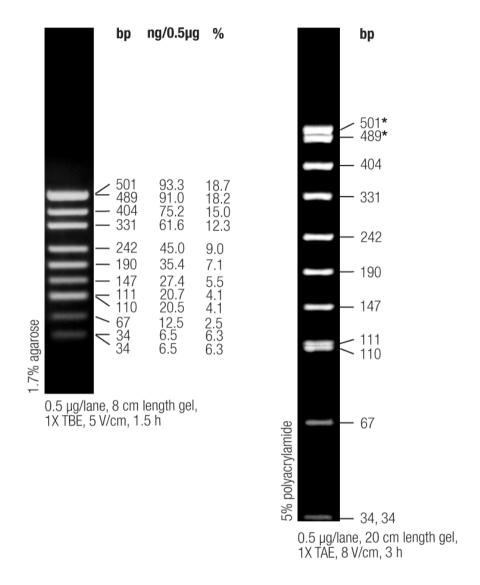
I. Loading on agarose gel:

- prepare the marker before loading:
 - 1 μ L (0.5 μ g) of the marker,
 - 1 μL of 6X DNA Loading Dye,
 - 4 μL of deionized water;
- vortex gently just prior to use;
- do not heat before loading;
- apply the prepared amount (6 μL) of the marker on a
 5 mm lane of agarose gel;
- following electrophoretic separation on gel, visualize the DNA bands by ethidium bromide staining.

Important Notes

- One vial (50 μg) is sufficient for ~100 applications.
- Use 0.1 μg (0.2 μL) of the marker (before dilution) per 1 mm of an agarose gel lane width.
- For DNA band visualization with SYBR® Green, GelRed and other intercalating dyes, do not add the dyes into the sample, use gel staining after electrophoresis or include dyes into agarose gel to avoid aberrant DNA migration.

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^{*}The 501 and 489 bp bands migrate anomalously (1, 2, 3). 26 bp fragment is not visible and comprises 1.0%

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II. Loading on polyacrylamide gel (1, 2, 3):

- prepare the marker before loading:
 - $2 \mu L (1 \mu g)$ of the marker,
 - 0.5 μL of 6X DNA Loading Dye,
 - 0.5 µL of deionized water;
- vortex gently just prior to use;
- do not heat before loading;
- apply the prepared amount (3 μL) of the marker on a 5 mm lane of polyacrylamide gel;
- following electrophoretic separation on gel, visualize the DNA bands by ethidium bromide staining.

Note

- One vial (50 μg) is sufficient for ~50 applications.
- Use 0.2 μg (0.4 μL) of the marker per 1 mm of a polyacrylamide gel lane width.

References

- 1. Stellwagen, N.C., Anomalous electrophoresis of deoxyribonucleic acid restriction fragments on polyacrylamide gels, Biochemistry, 22, 6186-6193, 1983.
- 2. Lane, D., et al., Use of gel ratardation to analyze protein nucleic acid interactions, Microbiological Reviews, 56, 509-528, 1992.
- 3. Stellwagen, N.C., Conformational isomers of curved DNA molecules can be observed by polyacrylamide gel electrophoresis, Electrophoresis, 21, 2327-2334, 2000.

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