

## PRODUCT INFORMATION

# AjuI

**#ER1951**      100 U

**Lot:** \_\_\_\_      **Expiry Date:** \_

5'...↓<sub>7</sub>(N)G A A(N)<sub>7</sub> T T G G(N)<sub>11</sub>↓...3'  
3'...↑<sub>12</sub>(N)C T T(N)<sub>7</sub> A A C C(N)<sub>6</sub>↑...5'

Concentration:      5 U/μL  
Source:              *Acinetobacter junii* RFL46  
Supplied with:      1 mL of 10X Buffer R  
                            1 mL of 10X Buffer Tango  
                            0.1 mL of 50X SAM Solution (0.5 mM)

**Store at -20°C**



BSA included

[www.thermoscientific.com/onebio](http://www.thermoscientific.com/onebio)

## RECOMMENDATIONS

**[1X Buffer R]+SAM** (for 100% AjuI digestion)

[10 mM Tris-HCl (pH 8.5 at 37°C), 10 mM MgCl<sub>2</sub>,  
100 mM KCl, 0.1 mg/mL BSA] +  
0.01 mM S-adenosylmethionine (SAM) (*see Note*).

### Incubation Temperature

37°C.

### Unit Definition

One unit is defined as the amount of AjuI at which no change in the fragmentation pattern is observed with further increase of enzyme. 1 μg of lambda DNA is incubated with the enzyme for 1 hour at 37°C in 50 μL of recommended reaction buffer.

### Dilution

Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/mL BSA and 50% glycerol.

### Double Digests

Thermo Scientific Tango Buffer is provided to simplify buffer selection for double digests. 98% of Thermo Scientific restriction enzymes are active in a 1X or 2X concentration of Tango™ Buffer. Please refer to [www.thermoscientific.com/doubledigest](http://www.thermoscientific.com/doubledigest) to choose the best buffer for your experiments.

1X Tango Buffer: 33 mM Tris-acetate (pH 7.9 at 37°C), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/mL BSA.

## Storage Buffer

Ajul is supplied in: 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM DTT, 1 mM EDTA, 0.2 mg/mL BSA and 50% glycerol.

## Recommended Protocol for Digestion

- Add:

nuclease-free water	16 $\mu$ L
10X Buffer R	2 $\mu$ L
DNA (0.5-1 $\mu$ g/ $\mu$ L)	1 $\mu$ L
50X SAM	0.4 $\mu$ L
Ajul	0.5-2 $\mu$ L
- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

The digestion reaction may be scaled either up or down.

## Recommended Protocol for Digestion of PCR Products Directly after Amplification

- Add:

PCR reaction mixture	10 $\mu$ L (~0.1-0.5 $\mu$ g of DNA)
nuclease-free water	18 $\mu$ L
10X Buffer R	2 $\mu$ L
50X SAM	0.6 $\mu$ L
Ajul	1-2 $\mu$ L
- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

## Thermal Inactivation

Ajul is inactivated by incubation at 65°C for 20 min.

## ENZYME PROPERTIES

### Enzyme Activity in Thermo Scientific REase Buffers, %

B <sub>+SAM</sub>	G <sub>+SAM</sub>	O <sub>+SAM</sub>	R <sub>+SAM</sub>	Tango <sub>+SAM</sub>	2X Tango <sub>+SAM</sub>
0-20	50-100	20-50	100	50-100	50-100

### Methylation Effects on Digestion

Dam: never overlaps – no effect.

Dcm: never overlaps – no effect.

CpG: may overlap – no effect.

EcoKI: never overlaps – no effect.

EcoBI: never overlaps – no effect.

### Stability during Prolonged Incubation

A minimum of 0.5 unit of the enzyme is required for digestion of 1  $\mu$ g of lambda DNA in 16 hours at 37°C.

### Digestion of Agarose-embedded DNA

A minimum of 5 units of the enzyme is required for digestion of 1  $\mu$ g of agarose-embedded lambda DNA in 16 hours.

### Number of Recognition Sites in DNA

$\lambda$	$\Phi$ X174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
3	1	0	0	0	0	0

### Note

- Ajul requires S-adenosylmethionine for activity.
- Complete cleavage of some substrates with Ajul is difficult to achieve.
- Ajul produces double-strand cuts on both sides of the interrupted recognition site. The exact cleavage position depends on the sequences flanking the recognition site and may shift by one base pair. However, for each individual sequence one cleavage position will dominate.

# CERTIFICATE OF ANALYSIS

## Overdigestion Assay

No detectable change in the specific fragmentation pattern is observed after a 160-fold overdigestion with Ajul (10 U/ $\mu$ g lambda DNA x 16 hours).

## Ligation and Recleavage (L/R) Assay

The ligation and recleavage assay was replaced with LO test after validating experiments showed LO test ability to trace nuclease and phosphatase activities with sensitivity that is higher than L/R by a factor of 100.

## Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded labeled oligonucleotides occurred during incubation with 10 units of Ajul for 4 hours.

## Blue/White (B/W) Cloning Assay

The B/W assay was replaced with LO test after validating experiments showed LO test ability to detect nuclease and phosphatase activities with sensitivity that equals to that of B/W test.

Quality authorized by:



Jurgita Zilinskiene

### **PRODUCT USE LIMITATION**

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Please refer to [www.thermoscientific.com/onebio](http://www.thermoscientific.com/onebio) for Material Safety Data Sheet of the product.

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